RECOMMENDED ORDER OF DRAW

POLICY STATEMENT

All PathWest Laboratory Medicine WA (LMWA) specimen collectors will collect blood sample tubes in a recommended order of draw to avoid cross contamination of additives between tubes (see Appendix A and B).

INTENT

To provide clarity and understanding to PathWest collection staff about the correct order of draw to be used when collecting blood samples while avoiding possible test error due to cross contamination of the sample by tube additives.

SCOPE

Appendix A of the policy applies to all PathWest staff performing phlebotomy collections. Appendix B is provided to external clients who request the PathWest policy be used in their clinical setting. Appendix C provides a combined tube guide and order of draw to ward staff. Appendices can be found at:

http://pathwest.health.wa.gov.au/HealthProfessionals/Pages/Collection-Guides.aspx

RESPONSIBILITIES

Employee

- Ensure they are familiar with the Order of Draw policy and how to access electronic or hardcopy information prior to performing specimen collection.
- To collect blood samples in the correct order prescribed by the Recommended Order of Draw policy to avoid cross contamination of additives between tubes.
- Read, sign and date document updates to acknowledge understanding of changes or amendments to the Order of Draw policy and forward acknowledgement to manager.

Pathology Specimen Collection Coordinator/Medical Scientist in Charge

- Ensure all staff have electronic or hardcopy access to the policy.
- Ensure all staff receive training and comply with the PathWest LMWA Recommended Order of Draw policy when collecting blood samples.
- Ensure the recommended order of draw is included in the staff annual competency assessment and records.
- Ensure all superseded documentation is replaced with the current version and staff are notified of any changes.

Document Author

 Undertake regular review, update documentation and maintain records of the notification date and the list of sites to which the documents are distributed.

Marketing

- Notify and inform external clients of changes to the policy including the distribution of updated documentation pertaining to the PathWest Recommended Order of Draw (Appendix A or Appendix C)
- Maintain register of external clients, including records for notification of updates i.e. date, reason for contact and name of PathWest staff member responsible for notifying clients.

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APPENDIX A PathWest Recommended Order of Draw

Recommended Order of Draw	Cap Colour(s)
Blood Cultures	
Sodium Citrate*	
Sodium Citrate (ESR tube) N.B can be collected at the end of the draw as tests performed on this tube are not compromised by other additives.	
ACD	
Serum with or without clot activator, with or without gel	
Rapid Serum Tube (RST) N.B. For use in Regional labs only	
Heparin with or without gel plasma separator	
QuantifFERON :TB Gold Plus Cellestis QuantiFERON In Tube vials	Order of collection for Quantiferon tubes: Grey, Green, Yellow Purple
EDTA with or without gel separator	
PTH (EDTA with gel)	
Trace metals EDTA If collecting for Chromium, Zinc and Cobalt, collect separately using a new luer adapter to avoid contamination	
Fluoride Oxalate	

^{*} Use of a winged collection set requires the collection of a discard tube if the first tube to be collected is a blue top Sodium Citrate tube. This discard tube will prime the tubing of the collection set and ensure that the correct anticoagulant/blood ratio is maintained. The discard tube should be a non-additive or light blue coloured tube and need not be filled.

Collection details for blood gas syringes (not part of the order of draw):

Lithium Heparin Gas Syringe

For arterial / venous blood gas testing Collect separately using a new luer adapter to avoid contamination. Transport at room temperature and test within 30 minutes of collection.



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APPENDIX B

PathWest Specimen Tube Collection Guide in Recommended Order of Draw

Notes

- 1. Blood gas syringes (arterial & venous) should be collected as per the procedure for Zinc collections.
- 2. **Haemolysis Warning** Never inject blood into the collection tube through a needle from a syringe.
- 3. Always mix the blood with anticoagulants by gently inverting the tube 6-8 times.
- 4. Do **not** shake tubes (exception is Quantiferon)
- 5. Do **not** top up or transfer blood from one tube to another.
- 6. Full patient identification is required for all pathology specimens (i.e.: full name and UMRN or full name and DOB)
- 7. **This is a guide only. For all Point of Care Tests,** or if you have any queries or require further information, please refer to the PathWest Test Directory or contact your local laboratory prior to sample collection.

TUBE CONTENT	VOLUME	TESTS / DETERMINATIONS	INSTRUCTIONS
Blood Cultures	8 -10 mls in each container	Blood cultures Investigation of a systemic bacterial or fungal infection	Must be collected using strict aseptic technique. First collect aerobic (grey/blue top) container followed by the anaerobic (purple/pink top)
Sodium Citrate	2.7 ml	INR, Prothrombin Time, APTT, Coagulation profile. (2.0 ml and 1.8 ml size tubes also available) NB. Please refer to your local testing laboratory for D-Dimer collection requirements.	Fill to the clear fill line marked on the tube. Send to the Lab within 4 hours of collection otherwise separate the centrifuged plasma within 4 hours of collection and transport chilled.
Sodium Citrate	1.36 ml	Erythrocyte Sedimentation Rate (ESR) (Note: 1.8 ml tubes are used at regional PathWest laboratories and cannot be forwarded to the QE11 Nedlands laboratory for testing)	Must fill tube to midway between marked lines N.B can be collected at the end of the draw as tests performed on this tube not compromised by other additives.
ACD	9.0 ml	HLA Typing – please check on line Specimen collection Manual before collection	Store at room temperature.

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Serum (+clot activator CAT)	6.0 ml	Note - a red top non-gel serum tube is preferred when collecting outside the metropolitan area. For collections in the metropolitan area please use a lithium heparin (dark green top – see below) Lithium assays can only be performed on a serum (red top) tube	Record the time of last dose on the request form and the collection date and time for all antibiotic assays & drug levels
Serum (+clot activator CAT)	6.0 ml	Vitamin B12, Serum Folate For collection requiring Thiamine or Red Cell Folate, additional samples will be required – please refer to EDTA	
OR Serum with Gel (+clot activator - SST II)	8.5 ml	Syphilis serology, Hepatitis serology, HIV, Viral serology, RRV, Arbovirus, CMV, Murray Valley Encephalitis, Q Fever, EBV, ANF, ASOT, RAST, Anti- DNaseB, TFT (T4 + TSH), PSA, Rheumatoid Factor (RF), , BHCG (Qualitative/Quantitative), Autoantibodies, Protein Electrophoresis, Most Hormones (FSH, LH, E2 etc), IM Screen (Monospot).	N/A
Rapid Serum Tube (RST)	6.0 ml	The RST tube is for HCG and Troponin when done in Regional labs and is required for PathWest's new biochemistry instrumentation.	N.B. RST tube may be used for chemistry and immunoassays at participating Regional sites. N.B. Not to be used for Drug levels
Lithium Heparin (LH)	4.0 ml	Therapeutic drug levels, antibiotic assays and drugs of abuse Note: A dark green top non-gel lithium heparin tube is preferred for samples collected within the metropolitan area. For samples collected outside the metropolitan area please use a serum (red top tube – see above) Do NOT collect Lithium Heparin (Green top) tubes for Lithium assays. Do NOT collect gel tubes for Troponin or D-Dimer assays Troponin I– Please collect a separate tube. Note:-proBNP– will also require a separate tube if tested in non-metropolitan laboratory.	Record the time of last dose on the request form and the collection date and time for all antibiotic assays & drug levels Do not spin and separate samples for Troponin, NT-proBNP or D-Dimer.
		NB. Please refer to your local testing laboratory for D-Dimer collection requirements.	

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Lithium Heparin (LH) OR Lithium Heparin Plasma with Gel (LH PST II)	8.0 ml	Biochemistry: UEC, LFT, Cardiac Enzymes, Lipids (Chol/Trig), HDL, Gamma GT, Calcium, Bicarbonate, Magnesium, Lipase, Uric Acid, Phosphate, CRP, Protein. NOTE: Iron Studies (Fe), Thyroid Studies (TFT), Hormones, PSA, Alcohol and some therapeutic drug levels can also be performed on this tube.	Calcium – Avoid venous occlusion during collection by releasing tourniquet ASAP. The UEC and Phosphate must be performed within 4 hours of collection otherwise separate the centrifuged plasma within 4 hours of collection then store and transport chilled. Unseparated samples should not be refrigerated but kept at room temperature until analysed.
EDTA (K2E)	4.0 ml	Haematology; FBC, Hb, Platelets, WCC, Reticulocytes, Glycated Hb (HbA1c), Malarial Parasites, G6PD Screen, Red Cell Folate, Cyclosporin, Hb electrophoresis. Heavy Metal Screen (2 tubes for mercury), FK506. Thiamine BNP will require EDTA plasma if tested at Nedlands. (Note Pro-BNP can be tested in some non-metropolitan laboratories. Please check with your local laboratory before collection).	Do not separate For Thiamine – please consult your local laboratory for collection requirements. Separate for BNP
or or	2 x 4.0 ml or 6.0 ml or 9.0 ml	Blood Group, Maternal antibody screen, Rhesus (Rh), Cross-match, Antibody Screen, Group & Hold, FMH, Kleihauer, Direct Coombs Test, Cord blood, PCR Studies	Specimen labels and request forms must have identical patient identification and include the collector's signature, date and time of collection. A formal declaration printed on the request form must be signed by the collector. Samples that do not meet these requirements will be discarded.
1111 121	4 x Cellestis QuantiFER ON In Tube vials	First-line Interferon-gamma release assay against tuberculosis-specific antigens, and used for the diagnosis of latent tuberculosis.	For each patient, collect 1ml of blood by venipuncture directly in to each of the QFT-Order of collection for Quantiferon tubes: Grey, Green, Yellow Purple

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EDTA Gel (K2E)	6.0 ml	Parathyroid Hormone (PTH) If an EDTA Gel tube is not available, please contact your local testing laboratory for further advice.	Do not confuse with PTH related protein (PTHrp) which requires a special tube. Please contact your local laboratory.
EDTA Trace Metal	6.0 ml	If an EDTA Trace Metal tube is not available, please contact your local testing laboratory for further advice. Zinc must be collected separately using a new luer adapter (see instructions) to reduce the risk of trace metal contamination from rubber bungs in other tubes. If other tubes are required the following instructions must be followed to avoid a double venepuncture:	For collections requiring a Trace Metal tube, please use a Winged Infusion Set (Butterfly Needle) and prepare a second (replacement) luer adapter. The first luer adapter must be changed immediately prior to collecting the Trace Metal tube to prevent contamination. Subsequent tubes can then continue to be collected as per the recommended order of draw. Please consult your local specimen collection team if assistance is required.
Fluoride	2.0 ml	Glucose only (Can be used for Glycated Hb)	Fill only to the mark. Do not separate.
Lithium Heparin Blood Gas Syringe	1.5 ml	Arterial / Venous Blood Gas testing.	Collect separately using a new luer adapter to avoid contamination. Transport at room temperature and test within 30 minutes of collection.

For all other tests or enquiries, please refer to your local PathWest laboratory or the online PathWest Specimen Collection Manual as special procedures may apply.

For transportation requirements, please refer to your local testing laboratory.

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APPENDIX C: Combined Tube Guide / Order of Draw for Ward Staff

APPENDIX C: Combined Tube Guide / Order of Draw for Ward Staff			
Hemogard™ Closure & Tube	TESTS & Information	SPECIAL INSTRUCTIONS	
Blood Cultures	Blood Culture	Fill the Aerobic Bottle [Blue] first, then the Anaerobic Bottle [Purple] Pink top is Pediatric Recommend at least 2 sets from a single site for routine collections, or 3 sets from 3 sites for ?Endocarditis	
Citrate	Coagulation Testing eg INR, APTT, Coag Screen, D Dimer, Protein C/S, ATIII NOTE: BLUE TOPS must be filled to the frosted band at the top of the tube or a recollection will be required.	Send to the lab within 4 hours of collection, or spin & separate plasma and transport chilled.	
Sodium Citrate [4NC]	ESR	FILL TO THE TOP	
ACD	HLA Studies		
Clot Tubes	Therapeutic Drug Levels [Use EDTA for Anti-Rejection Drugs]	NOTE Time & Date of Last Dose, Dosage and Collection Date & Time on the request form	
SST [with Gel]	All Referred Biochemistry [ie tests that take >1day for a result] All Viral Serology Immunology All Hormones		
RST [with Gel]	The RST tube is for HCG and Troponin when test is assayed in regional laboratories	N.B. RST tube may be used for chemistry and immunoassays at participating Regional sites. N.B. Not to be used for Drug levels	
Heparin	Troponin I & Creatinine [A&E iSTAT ONLY]	DO NOT SPIN DO NOT REFRIGERATE	
PST [with Gel]	Locally Performed Biochemistry [eg UEC, LFT, Lipids, CK, Lipase, CRP, Ca ²⁺ , Phos, Mg ²⁺ , LDH, NT proBNP]	Tests must be performed within 4 hours of collection for valid K^{\star} , or centrifuge tubes and store as per local guidelines."	
Cellestis QuantiFERON In Tube vials	First-line Interferon-gamma release assay against tuberculosis-specific antigens, and used for the diagnosis of latent tuberculosis	Order of collection for Quantiferon tubes: Grey, Green, Yellow Purple	
EDTA	Blood Count: FBC, FBP, CBC, WCC, Hb, Plt Glycated Hb: HbA ₁ C Malaria Blood Lead & Other Heavy Metals Anti Rejection Drugs [eg Tacrolimus, Sirolimus]	Do not separate	
or EDTA For crossmatch	Cross Matching and all related blood bank tests [eg Group & Hold, antibody screening, blood group] REMINDER: Minimum labelling for TM is FULL NAME & DOB, TIME & DATE of COLLECTION, and COLLECTORS SIGNATURE/ INITIALS on both form & tube. The signature/Initials on the tube and form must match	TUBE MUST BE SIGNED & HAVE TIME + DATE OF COLLECTION ON IT. UNSIGNED TUBES WILL BE DISCARDED AND A RECOLLECTION REQUIRED.	
Fluoride Oxalate	Blood Sugar	Fill to halfway mark	
Blood Gas Syringe	Venous or Arterial Blood Gases For venous /arterial blood gas testing.	Collect separately using a new luer adapter to avoid contamination. Transport at room temperature and test within 30 minutes of collection.	

SPECIMENS MUST BE <u>LABELLED</u> WITH PATIENTS SURNAME, FIRST NAME AND DATE OF BIRTH.

MIX <u>ALL</u> TUBES THOROUGHLY BY GENTLE INVERSION 5 – 6 TIMES.

FOR ANY OTHER TESTS PLEASE CONTACT THE LABORATORY

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